Chronic environmental exposure to lead affects semen quality in a Mexican men population

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Abstract

Background: Male infertility is affected by several factors. Lead is one of the heavy metals more bioavailable than usually modifies the sperm quality in humans.

Objective: The aim of this study was to establish the role of lead in semen quality in environmentally exposed men.

Materials and Methods: Semen and blood samples were obtained from two groups: the exposed group (EG=20) and the non-exposed group (NEG=27). Two semen aliquots were used, one to evaluate spermatic quality and the other for lead determination. Blood (PbB) and semen lead (PbS) determination was performed by atomic absorption spectrophotometry.

Results: The PbB concentration was significantly greater in the EG, 10.10±0.97 µgdL^-1 than in the NEG, 6.42±0.38 µgdL^-1 (p<0.01), as well as the PbS concentration, with 3.28±0.35 and 1.76±0.14µgdL^-1 in the EG and NEG respectively (p=0.043). A significant correlation between PbS and PbB concentration in the EG was found (r=0.573, p=0.038). Overall, the spermatic quality was lower in the EG than in the NEG. Specifically, there were significant differences in the spermatic concentration [EG=43.98±6.26 and NEG=68.78±8.51X10^6 cells.ml^-1 (p<0.01)], motility [EG=49±7 and NEG=67±4% (p=0.029)], viability [EG=36.32±3.59 and NEG=72.12±1.91% (p<0.01)] and abnormal morphology [EG=67±18 and NEG=32±12% (p<0.01)]. In the immature germ cells (IGC) concentration differences were found only for A cells (EG=8.1±1.1x100 and NEG=3.2±1.9X100 spermatozoa) (p<0.01) and for Sab cells (EG=3.4±2.2x100 and NEG=1.1±1.0X100 spermatozoa) (p=0.041).

Conclusion: These results suggest that chronic environmental exposure to levels of lead adversely affect the spermatic quality.

Key words: Lead, Spermatic quality, Infertility, Environmental.

Introduction

The infertility problems associated with lead in men environmentally and occupationally exposed have been reported (1-5). The findings of these prior studies are interesting but their results show variability in the association between lead and semen characteristics.

Working in smelters is an important environmental exposure factor for lead. Some authors have found that low or moderate seminal lead levels in smelter workers were correlated with a decrease in the semen quality; also, oligospermia, asthenospermia and an increased frequency of teratospermia have been reported in workers exposed to lead (2, 3). Some smelter workers were found to have hypogonadism and low serum testosterone concentration (1). On the other hand, men occupationally exposed to inorganic lead showed different endocrine and reproductive dysfunctions, particularly at the hypothalamic-pituitary-testicular axis (6-8). In this context, there are not published studies for Mexico that refer to the effects of lead on semen quality.

In order to establish the role of environmental lead exposure on semen quality (sperm quality is defined by the assessment of morphology, motility, viability and number of spermatozoa), we conducted this study in men living within a 500 meter radius of the third largest smelter and lead producer in the world, located in Torreón, State of Coahuila, México.
Materials and methods

This cross-sectional study was conducted in the Cellular Biology and Ultrastructure Department, Biomedical Research Center, School of Medicine of the Autonomous of Coahuila University. A total of 47 subjects were admitted for clinical and laboratory examinations. They were informed of the study objectives and given informed consent letters to sign (the protocol was approved by the Biomedical Research Center Committee, School of Medicine, University of Coahuila).

Inclusion criteria were age (18-30 years old), healthy, residents of the study area, similar social and economic status, 3 days of sexual abstinence. Exclusion criteria were subjects currently on any medication were not included. Also, patients with varicocele, leucospermia, smokers and alcoholic men were excluded from the study. The subjects of the study had lived in the same area for at least two years.

Twenty subjects who were residents within 500 meter radius from the metallurgic zone formed the exposed group (EG: n=20), and 27 subjects residing in two communities at a distance of 15 and 18 km, respectively from this zone formed the non-exposed group (NEG). In order to control for possible confounding sources of lead, all subjects were interviewed using a questionnaire, which included items on living conditions, socioeconomic status and occupational factors.

Sample collection and lead assay

The semen samples were collected by masturbation after at least 4 days of abstinence. The materials used for sample collection were plastic containers (125 ml), Falcon tubes (conical polystyrene, 15 ml) and polyethylene tubes (5 ml) (Becton Dickinson Labware, USA). These were cleaned with 10% nitric acid and rinsed with deionized water. The semen specimens were placed in an incubator at 37°C immediately after collection.

After liquefaction, ejaculates were divided into two aliquots. One aliquot was used to carry out routine laboratory analysis: seminal volume, sperm count, total amount of sperm per ejaculate, sperm motility and viability in accordance with the guidelines recommended by the World Health Organization. Smears for IGC and spermatoc morphology determinations were stained with the modified Bryan/Leishman stain (9). The second semen aliquot was used for lead determination. Lead analysis in blood and semen samples were measured by atomic absorption spectrophotometry with a heated graphite furnace according to Brown et al (10).

Statistical analysis

The seminal sperm concentration, progressive sperm motility, percentage of viable sperm and sperm morphology were analyzed by analysis of variance (ANOVA). Immature germ cells (IGC) are considered all round cells not identified as leukocytes, these cells are: spermatogonia, primary and secondary spermatocites and Sab, Scd spermatids. The germinal cell classifications are based on those proposed by Macleod with modifications by WHO (9). The IGC concentration per field was calculated as follows (11):

\[
\text{Concentration} = \frac{\text{Number of IGC}(\text{Sperm Count/mL})}{100\text{Spermatozoa}}
\]

Plasmatic and seminal lead concentrations were analyzed by ANOVA between groups and by simple correlation. Regression analysis was used to assess the association between lead semen concentration and spermatoc quality in the EG. The results are presented as the mean with standard error of mean (SEM). The STATA version 6 program was used for all statistical analysis (12).

Results

The average age in the EG was 25±1.30 years (range 22-27) and the NEG was 25±1.24 years (range 20-26). Semen lead levels in all subjects were always less than this level in blood and were significantly higher in EG (3.28±0.35 nmol/L) in comparison to NEG (1.76±0.14 nmol/L), p=0.043 (Table 1).

The mean blood lead concentration in the EG was 10.10±0.97 nmol/L and for NEG 6.42±0.38 nmol/L (p<0.01). A significant correlation between the levels of lead in semen and in blood of the EG were observed (r=0.573, p=0.038). No significant correlations (p=0.071) were found between lead semen concentration and seminal parameters (vs.
spermatic concentration \((X 10^6 \text{ cell/mL})\): 
\(r=-0.006;\) vs. spermatic viability: 
\(r=0.244\) and vs. spermatic motility: 
\(r=-0.05\). Semen quality parameters in men was affected in all parameters for both groups (Table II).

A significant higher percentage \((p=0.029)\) of mobile spermatozoa were found in NEG \((67\pm4\%)\) as compared to EG \((49\pm7\%)\). The mean sperm concentration was also significantly higher \((p<0.01)\) in NEG than EG, 68.78\(\pm\)8.51 and 43.98\(\pm\)6.26\(\times\)10^6 \text{ cell/mL}, respectively. The volume was similar in the NEG \((2.9\pm0.26 \text{ mL})\), compared to EG \((3.2\pm0.32 \text{ mL})\). On the other hand, an increased percentage of morphologic abnormalities were observed in EG \((67\pm18\%)\) compared to 32\(\pm\)12\% in NEG \((p<0.01)\) (Table III).

The predominant abnormalities were defects of the middle piece, including cytoplasmatic droplets \((41\%)\), and defects of the head \((\text{large oval head} (15\%), \text{pyriform head} (3\%) \text{and duplicate head} (5\%)\) (Figure 1). Significant differences were found between the percentage of germinal cells secretion observed in the EG in comparison with NEG \((\text{global mean by germinal cell by group}: p<0.01)\). The principal germinal cells secretion were spermatogonia type A, and germinal cells as Sab and Scd (Table IV, Figure 1).

### Table I. Blood and semen lead concentration in the two study groups (Mean±SEM)

<table>
<thead>
<tr>
<th>Sample</th>
<th>Lead concentration (nmol/L)</th>
<th>Range</th>
<th>Range</th>
</tr>
</thead>
<tbody>
<tr>
<td>Blood</td>
<td>10.10 (\pm) 0.97**</td>
<td>(4.4-22.7)</td>
<td>6.42 (\pm) 0.38</td>
</tr>
<tr>
<td>Semen</td>
<td>3.28 (\pm) 0.35*</td>
<td>(0.86-7.44)</td>
<td>1.76 (\pm) 0.14</td>
</tr>
</tbody>
</table>

\(**p<0.01, *p<0.05\)

EG: exposed group, NEG: non-exposed group.

### Table II. Seminal quality in the study groups (Mean±SEM)

<table>
<thead>
<tr>
<th>Seminal parameters</th>
<th>NEG</th>
<th>EG</th>
</tr>
</thead>
<tbody>
<tr>
<td>Concentration ((X 10^6\text{cells/mL}))</td>
<td>68.78 (\pm) 8.51**</td>
<td>43.98 (\pm) 6.26</td>
</tr>
<tr>
<td>Motility (%) a</td>
<td>67 (\pm) 4*</td>
<td>49 (\pm) 7</td>
</tr>
<tr>
<td>Viability (%) b</td>
<td>72.12 (\pm) 1.9*</td>
<td>36.32 (\pm) 3.59</td>
</tr>
<tr>
<td>Morphology (%) c</td>
<td>33 (\pm) 12</td>
<td>68 (\pm) 18**</td>
</tr>
<tr>
<td>Volume (ml)</td>
<td>2.9 (\pm) 0.26</td>
<td>3.2 (\pm) 0.32</td>
</tr>
</tbody>
</table>

a) Show total motility percent by group.
b) Show the live motile and live immotile spermatozoa percent by group.
c) Show normal morphology percent by groups.

\(^*p<0.05, **p<0.01\)

EG: exposed group, NEG: non-exposed group.

### Table III. Spermatic morphology determination in the study groups (Mean±SEM)

<table>
<thead>
<tr>
<th>Form</th>
<th>NEG (%)</th>
<th>EG (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Normal</td>
<td>68**</td>
<td>33</td>
</tr>
<tr>
<td>Large oval head</td>
<td>11</td>
<td>18*</td>
</tr>
<tr>
<td>Small oval head</td>
<td>8**</td>
<td>1</td>
</tr>
<tr>
<td>Amorphous middle piece</td>
<td>11</td>
<td>41**</td>
</tr>
<tr>
<td>Duplicate head</td>
<td>1</td>
<td>5*</td>
</tr>
<tr>
<td>Duplicate tail</td>
<td>1</td>
<td>2</td>
</tr>
</tbody>
</table>

\(^*p<0.05; **p<0.01\)

EG: exposed group, NEG: non-exposed group.
Table IV. Principal immature germ cells concentration in the study groups (Mean±SEM)

<table>
<thead>
<tr>
<th>Cell Type</th>
<th>NEG (X100 spermatozoa)</th>
<th>EG (X100 spermatozoa)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Spermatogonia A</td>
<td>3.2 ± 2</td>
<td>8.1 ± 1 **</td>
</tr>
<tr>
<td>Spermatid Sab</td>
<td>1.1 ± 1</td>
<td>3.4 ± 2 *</td>
</tr>
<tr>
<td>Spermatid Scd</td>
<td>2.1 ± 1</td>
<td>2.8 ± 2</td>
</tr>
</tbody>
</table>

*p<0.05, ** p <0.01
EG: exposed group, NEG: non-exposed group.

Discussion

The results of our study show that exposure to lead is related to decreased of the quality characteristics in semen in environmental exposed men. In this study, the EG lead blood concentration was higher than NEG. Blood studies made in the same smelter exposed area, show the results in the adult exposed group and control group were 19.2±4.5 and 8.9±1.3nmol/L (p<0.001) respectively, using the same method for lead determination used in this study. On the other hand, in children (7-12 years old) exposed environmentally to lead in the same area a significant difference was found when compared with a control group (17.3±56 and 6.4±3.1 nmol/L respectively) (p<0.01) (13, 14). Garcia-Vargas et al showed results in children living nearest school refinery with 27.6 nmol/L mean of lead blood (15).

These studies show similar results in lead blood and suggest a potential risk of exposition of people, mainly of young and mature men, and its repercussion in health, showing the latent environmental chronic exposure in this population (13-15). On the other hand, our results show a decrease in the seminal parameters in the exposed men for spermatic concentration, asthenospermia and
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Some actions of lead on the male reproductive system of different species includes infertility, germinal epithelial damage, oligospermia and testicular degeneration, decreased sperm motility, prostate hyperplasia, abnormal sperms, teratospermia, hypospermia and asthenospermia (29). As other heavy metals, lead seems to exert its toxicity upon different subpopulations of testicular cells. Mechanisms of heavy metal toxicity vary and not only include different cell sensitivities, but also direct vs. indirect actions.

Furthermore, it appears that primary damage to one cell type may secondarily affect other cell types in the testes. For example, in the case of lead, there are direct evidences for testicular toxicity in germinal cells and Leydig cells, but, for Sertoli cells toxicity is possible a secondary effect. The cellular motility in the present study was different for the NEG and the EG (67±4 and 49±7% respectively) (p=0.029), similar results were found in infertile men for this parameter by El-Zohairay et al (33.00±5.8%) and Saaranen et al (43.9±18.9%) (15, 18).

However, in our work, a positive correlation was not found between lead in semen concentration and seminal parameters. Similar results were found by Saaranen et al, Jockenhövel et al and Noack-Füller et al (6, 16, 30). The smelter has been contaminating this area for more than a hundred years (starting in 1887), it is the first silver producer in the world (year 2003: 78, 575, 158 Oz), and the first zinc (year 2003: 209, 403 tons), lead (year 2003: 125, 735 tons), and gold (year 2003: 1, 056, 385 Oz) producer in Latin America.

In the studied area, the bioavailability (air) and accumulative availability (dust and soil) of lead concentrations are critical. For example, the lead concentration in total air suspended particles (TSP) and suspended particles of size <10μm (PM-10) were 6.1 and 3.8 μg/m³ in a schools near the smelter zone in the same area of our study (15). These values are higher than the parameters suggested by the U.S. National Ambient Air Quality Standard, air values used in Mexico are 1.5 μg Pb/m³ (31). In soil and dust, the main problem was in the superficial layers (same on schools), the results show in soil a concentration of 2772.8 μg Pb/g (layer 0-10 cm), 2374.7μg Pb/g (layer 10-20 cm) and dust 3471.3μg Pb/g, these values surpassed the U.S. Environmental Protection Agency parameters (200-500 μg/g soil) (16, 32).

The environmental exposure in this smelter complex shows a potential and different contamination manner in this population. Allowed concentrations of toxic contaminants are still a debatable issue in developing countries, where there is wide variation in human susceptibility and tolerance, depending on general human health and other environmental factors. Therefore, this chronic exposure on this population in this risk area is critical and there should be other control and prevention strategies.

**Conclusion**

In conclusion, this study shows that the environmental exposure to low levels of lead affect the spermatic quality in an impairing way. Other studies about occupational and environmental lead exposure effects and spermatic quality in this risk area would be very interesting.

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**Conflict of interest**

The authors indicate no potential conflicts of interest.
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References


12. STATA version 6. Stata Corporation, College Station, TX, USA.


29. Bell JU, Thomas JA. Effects of lead on the reproductive system. In Singhal RL, Thomas JA

